

PHASE SEPERATION IN ARTIFICIAL CELLS OF POLY (2-OXAZOLINE) AND POLYLACTIDE COPOLYMERS



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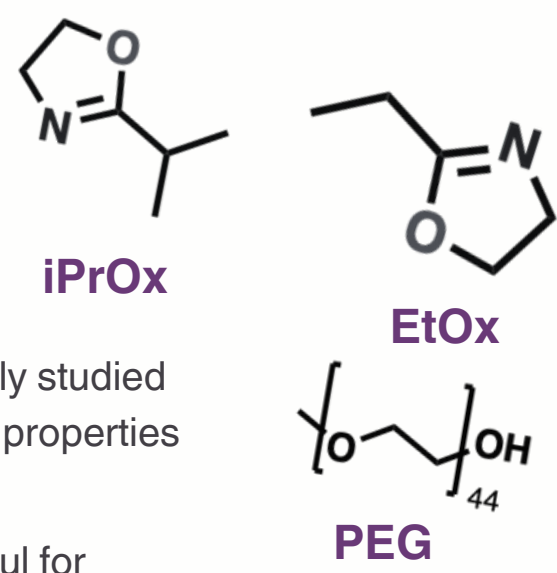
Introduction

Phase-separated artificial cells can provide valuable insight into the fundamental processes of natural cells by mimicking their properties and functionalities.¹

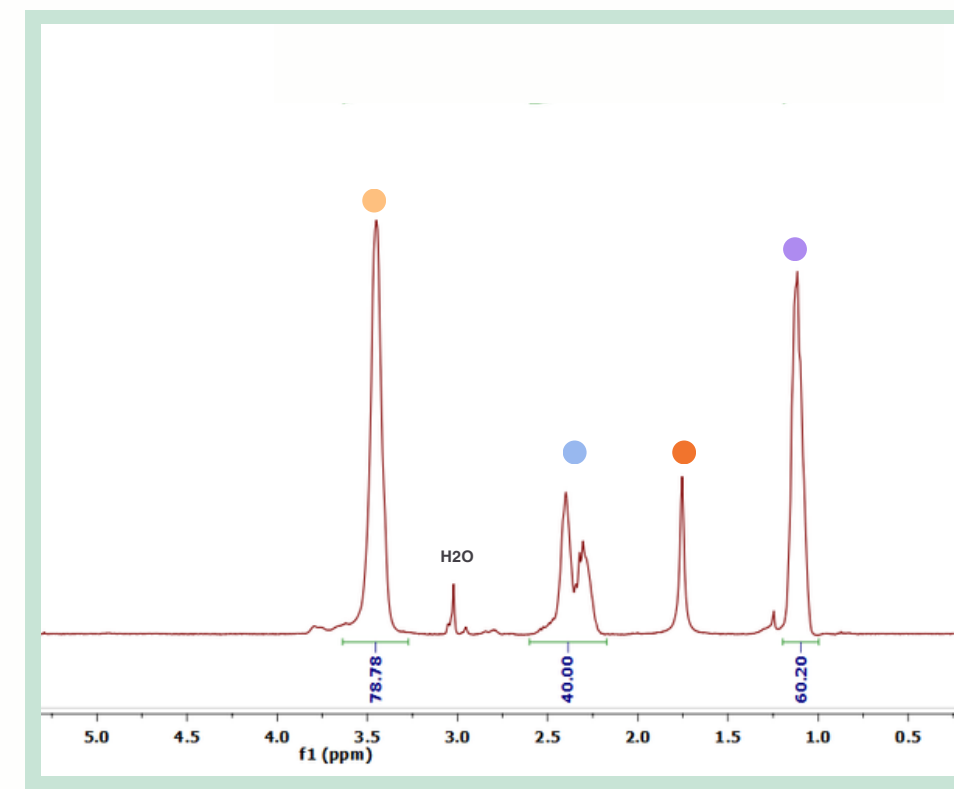
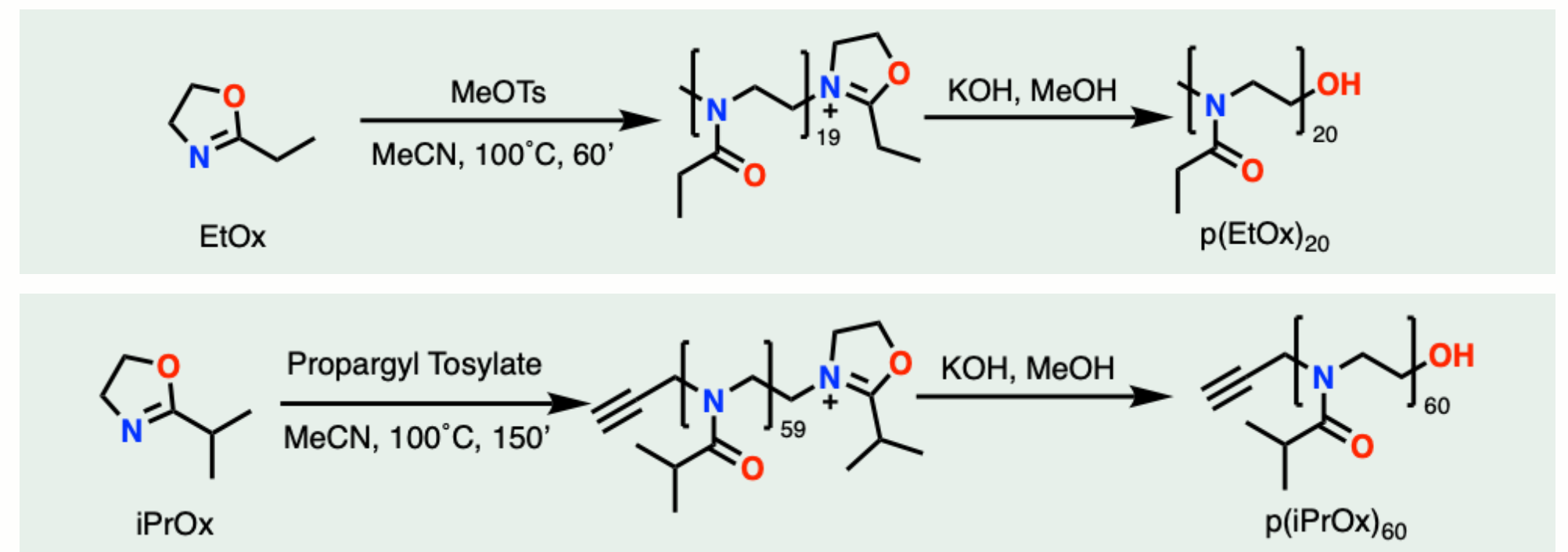
Poly(2-oxazolines) are a promising class of polymers that have been extensively studied due to their potential for a wide range of applications.² They possess desirable properties such as water

solubility, low cytotoxicity and stealth effects which make them particularly useful for nanomedicines and drug delivery.³ It is also known that poly D,L-lactides are highly biocompatible due to low toxicity and easy metabolism.² The synthesised amphiphilic block copolymers can be used as model membranes to explore the behaviours of biomolecules.¹ These polymers have gained particular interest in the biomedical field due to their ability to form nanostructures.⁴

The aim: Synthesising poly(2-ethyl-2-oxazoline)-*b*-PDLLA, poly(2-isopropyl-2-oxazoline)-*b*-PDLLA and PEG-*b*-PDLLA based block copolymers, using ring-opening polymerisation (ROP) to formulate phase-separated Giant Unilamellar Vesicles (GUVs).



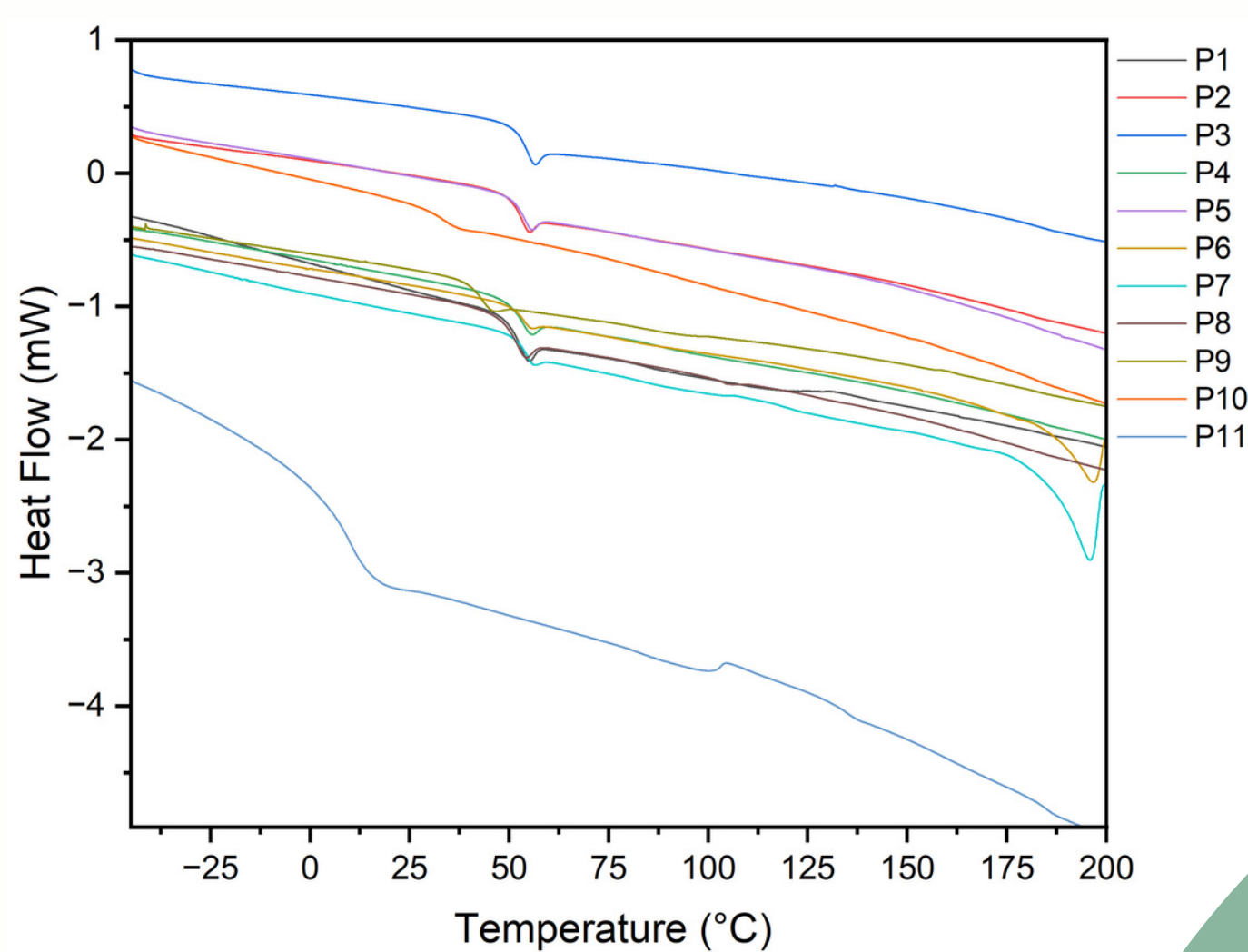
Cationic Ring-Opening Polymerisation of Macroinitiators



- CROP was used to form the homo-polymers EtOx and iPrOx, varying the initiators (methyl or propargyl tosylate)
- All eight homopolymers had a polydispersity of $\bar{D} \leq 1.21$

Table 1. Molecular weight data of all synthesised homopolymers.

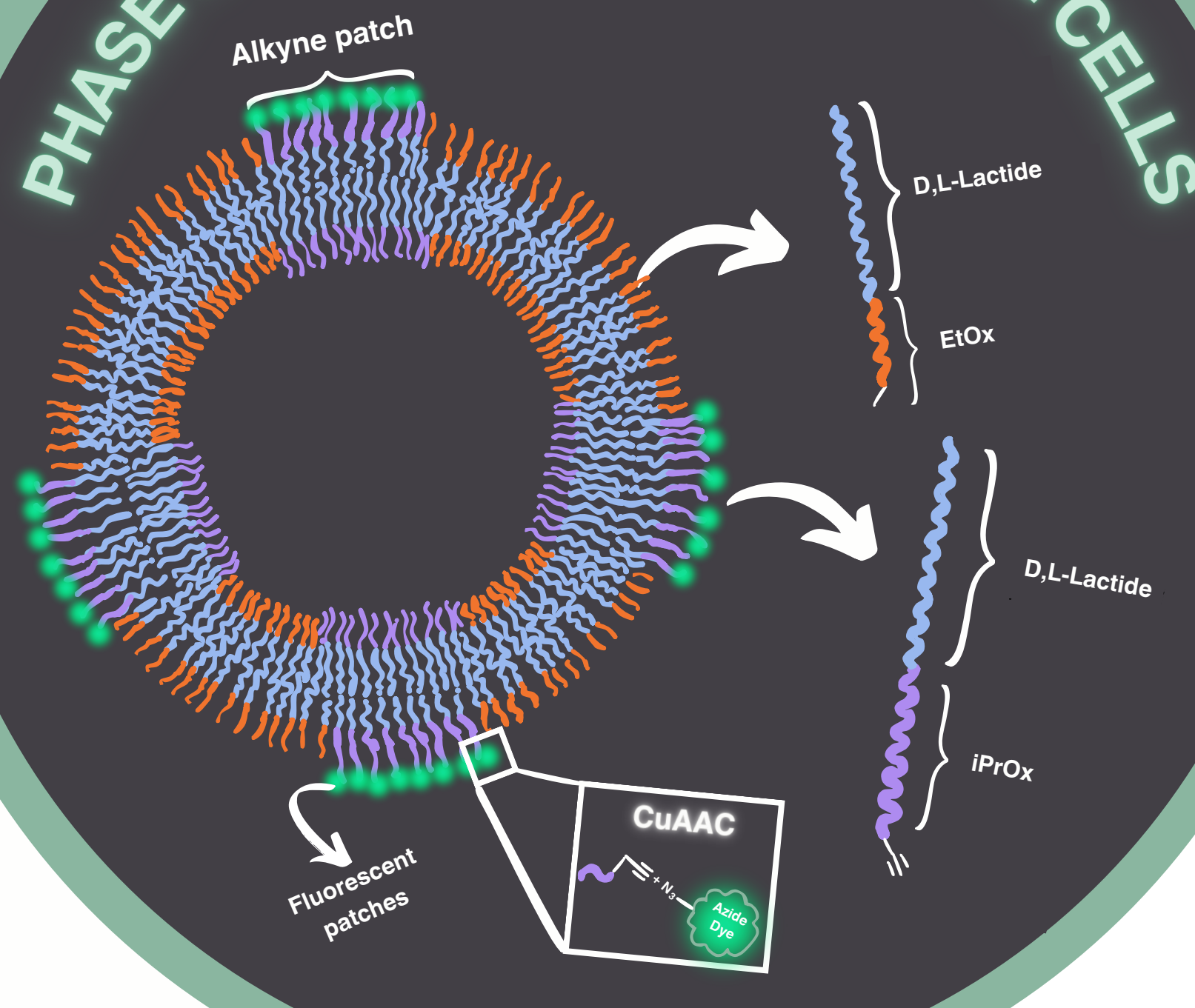
| Polymer | M _n , NMR (Da) | M _n , GPC (Da) | \bar{D} |
|-------------------------------|---------------------------|---------------------------|-----------|
| Methyl-EtOx ₂₀ | 1900 | 2000 | 1.12 |
| Propargyl-EtOx ₂₀ | 2000 | 1900 | 1.20 |
| Methyl-EtOx ₆₀ | 6000 | 5900 | 1.19 |
| Propargyl-EtOx ₆₀ | 6000 | 5800 | 1.21 |
| Methyl-iPrOx ₂₀ | 2100 | 2600 | 1.11 |
| Propargyl-iPrOx ₂₀ | 2000 | 2100 | 1.19 |
| Propargyl-iPrOx ₆₀ | 6000 | 5700 | 1.13 |
| Methyl-iPrOx ₆₀ | 5900 | 7300 | 1.12 |



Differential Scanning Calorimetry

The T_gs ranged from 52 to 55 °C (EtOx/iPrOx-*b*-PDLLA). The T_gs decreased as PEG length within the diblock increased (for P9 to P11 they were 43, 33 and 10 °C, respectively).

PHASE SEPERATION IN ARTIFICIAL CELLS



Diblock copolymer synthesis

The synthesised polyoxazolines were the macroinitiators for ring-opening polymerisation (ROP) of D,L-lactide. The DP of the hydrophobic D,L-lactide block was targeted to be 120 (targeted M_n = 17 kDa).

Table 2. Molecular weight data of polymers P1-P11.

| Polymer | Composition | M _n , NMR (Da) | M _n , Theo (Da) | M _n , GPC (Da) | \bar{D} |
|---------|---|---------------------------|----------------------------|---------------------------|-----------|
| P1 | p(EtOx) ₁₉ - <i>b</i> -PDLLA ₁₂₃ | 17700 | 19600 | 14100 | 1.12 |
| P2 | p(iPrOx) ₂₀ - <i>b</i> -PDLLA ₁₂₈ | 18500 | 20500 | 14500 | 1.26 |
| P3 | p(iPrOx) ₂₁ - <i>b</i> -PDLLA ₁₂₄ | 17900 | 20000 | 12200 | 1.17 |
| P4 | p(EtOx) ₆₀ - <i>b</i> -PDLLA ₁₁₅ | 16600 | 22600 | 12500 | 1.26 |
| P5 | p(EtOx) ₅₉ - <i>b</i> -PDLLA ₁₃₁ | 18900 | 19500 | 15600 | 1.27 |
| P6 | p(iPrOx) ₅₉ - <i>b</i> -PDLLA ₁₁₃ | 17500 | 23400 | 15500 | 1.27 |
| P7 | p(iPrOx) ₆₀ - <i>b</i> -PDLLA ₁₂₁ | 16000 | 22000 | 20500 | 1.28 |
| P8 | p(EtOx) ₆₀ - <i>b</i> -PDLLA ₁₁₁ | 17500 | 16200 | 13000 | 1.31 |
| P9 | PEG ₁₇ - <i>b</i> -PDLLA ₁₂₁ | 17300 | 17400 | 13500 | 1.24 |
| P10 | PEG ₄₄ - <i>b</i> -PDLLA ₁₂₀ | 17300 | 17400 | 21200 | 1.17 |
| P11 | PEG ₁₁₅ - <i>b</i> -PDLLA ₁₁₆ | 16700 | 20500 | 17100 | 1.31 |



- Copolymers have polydispersities of $\bar{D} \leq 1.31$
- M_n of lactide block calculated by ¹H-NMR (Table 2, M_{n,NMR})

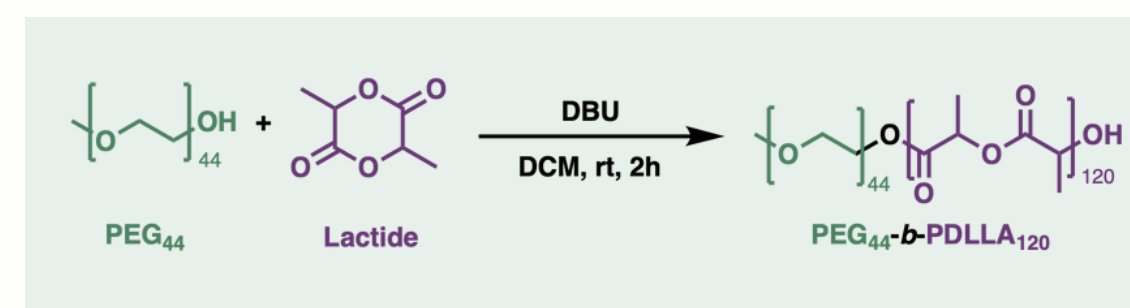


Figure 5. GPC traces of the homopolymer and resulting diblock copolymer. Measurements were performed using THF (2% TEA and 0.01% BHT) as the eluent. PMMA standards were used for the calibration.

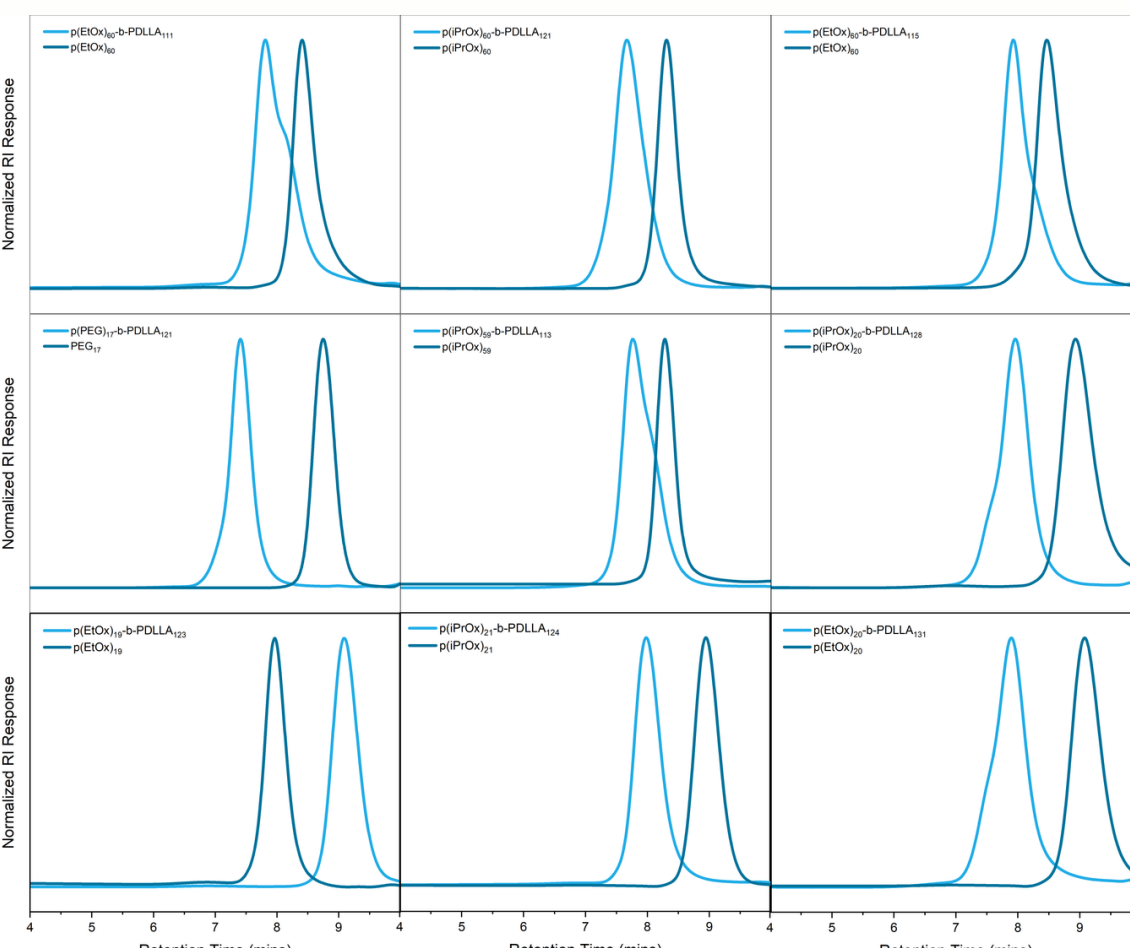
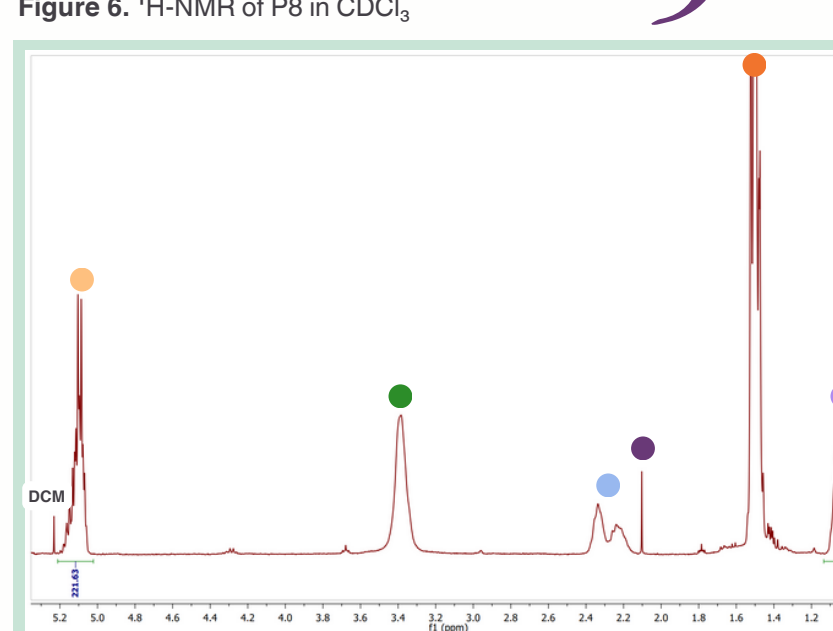


Figure 6. ¹H-NMR of P8 in CDCl₃



- ¹H-NMR analysis (Figure 6) shows expected product peaks for the ring opening of D,L-lactide with an oxazoline macroinitiator.
- The GPC shifts for the copolymers compared to the homopolymers (Figure 5) show the evolution of molecular weight.

Kinetics

Living polymerization was confirmed by kinetic analysis of the homopolymerizations (Figure 2,4), showing that each initiator initiated a monomer chain. The trend in Figure 4 shows the GPC traces shift to the left as the homopolymerisation occurs, depicting a clear evolution of molecular weight.

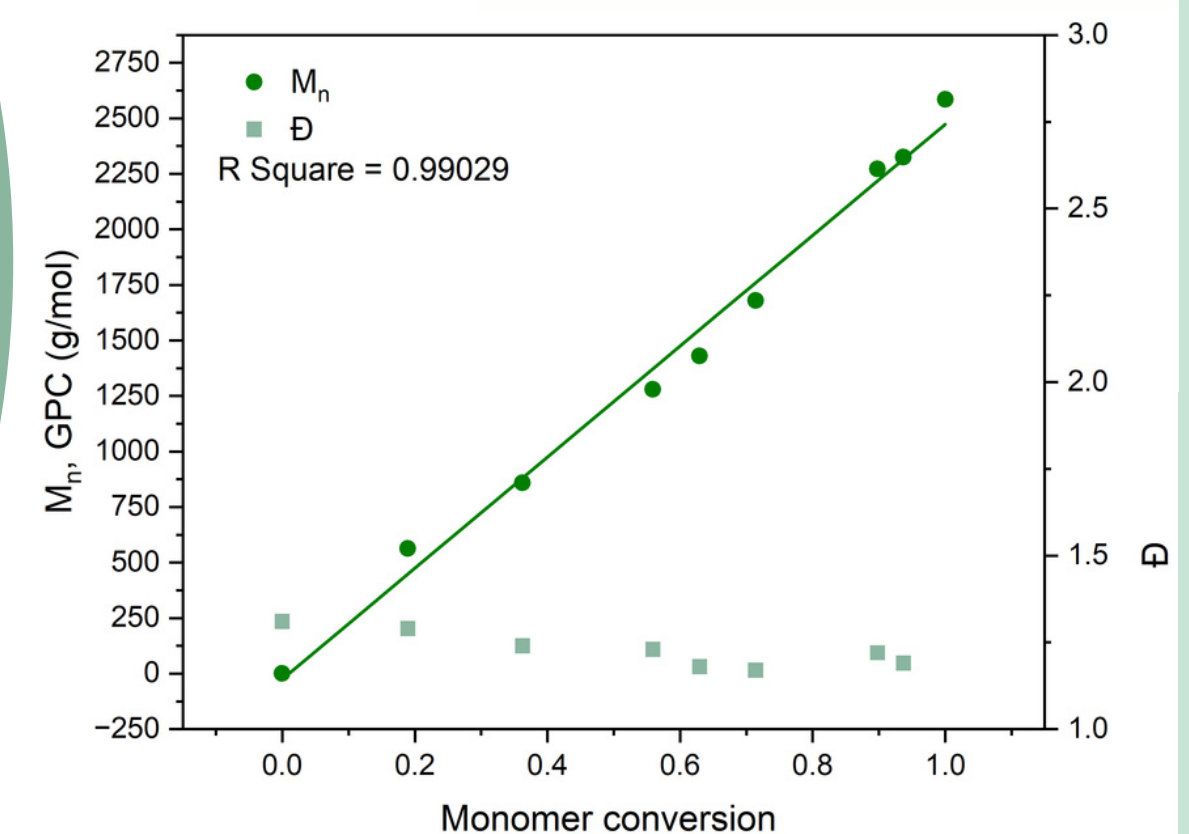


Figure 2. Graph showing monomer conversion against M_n, GPC and the polydispersity values.

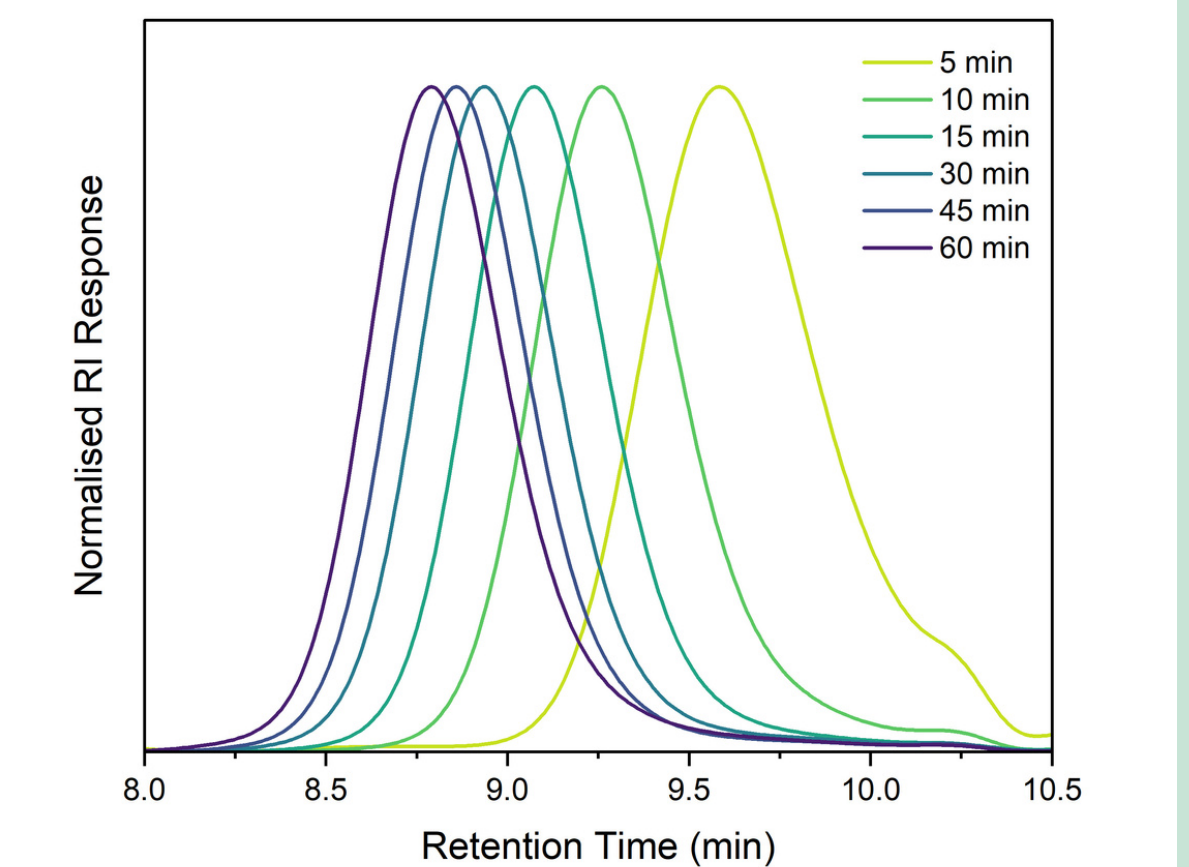
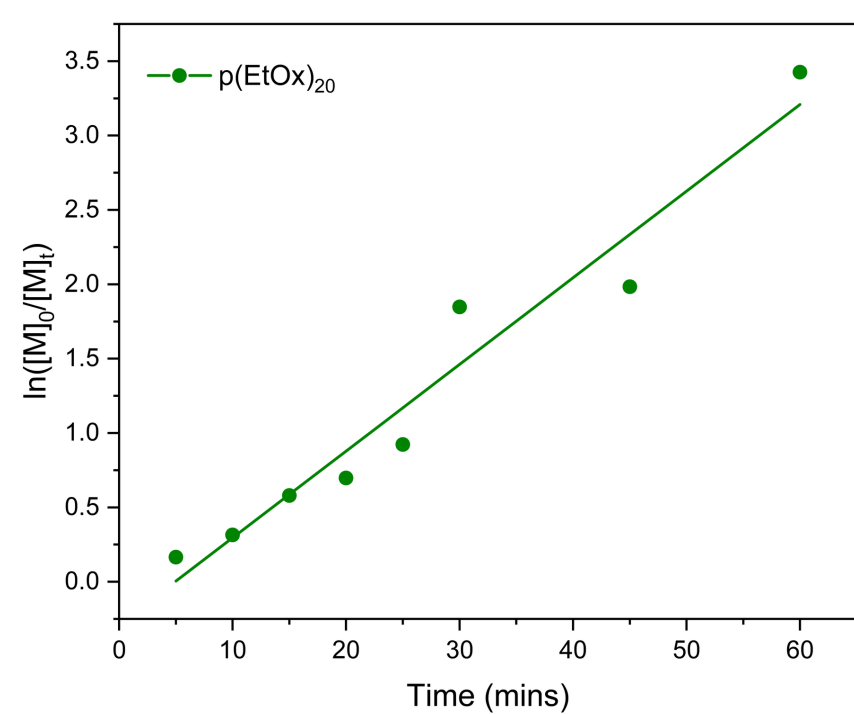


Figure 3. GPC traces of the homopolymer and resulting diblock copolymer. Measurements were performed using THF (2% TEA and 0.01% BHT) as the eluent. PMMA standards were used for the calibration.

Figure 4. Graph showing monomer conversion against M_n, GPC and the polydispersity values.



Conclusion and future work

In summary, a series of the homopolymers 2-ethyl-2-oxazoline and 2-isopropyl-2-oxazoline followed by eleven diblock copolymers of D,L-lactide were synthesised. The ROP procedure yielded well-defined and narrowly dispersed copolymers.

In collaboration with Prof. Sébastien Lecommandoux's group at LCPO in Bordeaux, future work includes the study of phase-separated membranes by co-formulating Giant Unilamellar Vesicles (GUVs) by varying the ratio between the synthesised diblock copolymers. Localisation of alkyne groups would be proven by labelling the alkyne functionality groups with azide dyes.

References

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